Lesson plan

Name of the Faculty : Ms Pratima Saini

Discipline : DMLT

Semester : 3rd

Subject : Clinical Microbiology III

Lession Plan Duration: 15 weeks (from October, 2021 to January, 2022)

Work load (Lecture / practical) per week (n hours) = Lecture=3, Practical=6

WORK	THEORY		Practical	
	Lecture Day	**Topic (Including assignment/test}**	**Practical Day**	**Topic**
1st	1	Introduction to medical parasitology	L1	Collection and routine stool examination for detection of intestinal parasites by Saline preparation
	2	General characteristics, morphology of Protozoa		
	3	Classification of Protozoa		
2nd	4	General characteristics, morphology of Helminthes	L2	Collection and routine stool examination for detection of intestinal parasites by Iodine preparation
	5	classification of Helminthes		
	6	Collection, transportation, processing and preservation of blood sample for routine investigations		
3rd	7	Collection, transportation, processing and preservation of stool sample for routine investigations	L3	Collection and routine stool examination for detection of intestinal parasites by Floatation method (saturated salt solution/zinc sulphate)
	8	Introduction about Concentration techniques		
	9	- Principle and application of concentration techniques(floating techniques)		
4th	10	Simple floating technique	L4	Collection and routine stool examination for detection of intestinal parasites by Centrifugation method (formal ether)
	11	DCF technique		
	12	Sedimentation techniques(simple)		
5th	13	Sedimentation techniques(formalin ether)	L5	Identification of Tapeworm from preserved specimen/slides
	14	Introduction about Giardia and Morphology of Giardia		

	15	Life cycle and Lab diagnosis of Giardia		
6th	16	Morphlogy and Life cycle of Entamoeba histolytica	L6	Identification of Roundworm from preserved specimen/slides
	17	Lab diagnosis of Entamoeba histolytica		
	18	Morphology and Life cycle of Ancylostoma		
7th	19	Life cycle and Lab diagnosis of Ancylostoma	L7	Identification of Hookworm from preserved specimen/slides
	20	Morphology of Ascaris lumbricoides		
	21	Life cycle and Lab diagnosis of Ascaris lumbricoides		
8th	22	Assignment	L8	Identification of Pinworm from preserved specimen/slides
	23	Morphology and life cycle of . T solium,		
	24	Lab diagnosis T solium,		
9th	25	Morphology and life cycle of T saginata	L9	Identification of Trichamonas vaginilis from preserved specimen/slides
	26	Lab diagnosis T saginata		
	27	Introduction about Malarial Parasite		
10th	28	Morphology of P. Vivax	L10	Preparation of stains (Leishman, Giemsa, Field)
	29	Life cycle of P. Vivax		
	30	Lab diagnosis of P. Vivax		
11th	31	Morphology of P. Falciparum	L11	Preparation of thin and thick smears
	32	Life cycle of P. Falciparum		
	33	Lab diagnosis of P. Falciparum		
12th	34	Assignment	L12	Staining of smears by Leishman, Giemsa, Field
	35	Introduction about Virology		
	36	General Characteristics of virus		
13th	37	Structure of viruses.	L13	Examination of smears for malarial parasite (P. vivax)
	38	Classification of virus		
	39	Lab diagnosis and prevention of – - Rabies - Polio		
14th	40	Lab diagnosis and prevention of – HIV , HBV (Hepatitis 'B' virus)	L14	Examination of smears for malarial parasite(P. falciparum)
	41	Introduction about Virological Samples		

		- Collection of Virological Samples - Transportation - Storage		
	42	Transportation of virological samples		
15th	43	Storage of virological samples	L15	Demonstration of various stages of malarial parasite from stained slides
	44	Assignment		
	45	Test		